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Synthesis of protected derivatives and short peptides of antAib, a novel C^{α} -tetrasubstituted α -amino acid of the Ac₅c type possessing a fused anthracene fluorophore

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Abstract—The N^{α} -Boc and N^{α} -Fmoc protected derivatives of 2-amino-2,3-dihydro-1*H*-cyclopenta[*b*]anthracene-2-carboxylic acid (antAib), a novel fluorescent, achiral, α -amino acid, rigid analogue of the known 9-antAla and 2-antAla residues, and belonging to the class of $C_i^{\alpha} \rightarrow C_i^{\alpha}$ cyclized, $C^{\alpha,\alpha}$ -disubstituted glycines (strong β -turn and helix inducers in peptides), were synthesized in seven steps from 1,2,4-trimethylbenzene. The UV absorption and fluorescence properties of Boc–antAib–OEt and Boc–antAib–OH are also described. Solution syntheses of the short peptides Boc–antAib–L-Ala–OMe, Fmoc–L-Ala–antAib–L-Ala–OMe, as well as Boc–Aib–antAib–L-Ala–OMe and the side product 2,5-dioxopiperazine *cyclo*-[antAib–L-Ala], are presented as examples of the coupling ability at both C- and N-termini of the antAib residue. © 2006 Elsevier Ltd. All rights reserved.

1. Introduction

Fluorescence spectroscopy has become a highly valuable technique for conformational studies of biopolymers, the development of peptide-based chemosensors, and biochemical research in general.¹ Incorporation of a fluorescent probe into a peptide chain may be achieved by reaction with side-chain functional groups or the direct use of fluorophore-bearing amino acids. In this connection, *synthetic* fluorescent amino acids may exhibit significant advantages over the related *protein* (Trp, Tyr) residues in terms of potentially different and improved properties. In previous studies, we took advantage of the fluorescence, the increased rigidity, and the axial chirality of 2',1':1,2;1'',2'':3,4-dinaphthcyclohepta-1,3-diene-6-amino-6-carboxylic acid (Bin)² (Fig. 1),

a $C^{\alpha,\alpha}$ -disubstituted glycine derived from 1,1'-binaphthyl, to carry out photophysical studies involving intramolecular energy transfer (fluorescence quenching) and intramolecular spin polarization (CIDEP) effects in conformationally constrained peptide-based systems.³ However, interpretation of the data was complicated by the nonplanar structure of the Bin 1,1'-binaphthyl core. To circumvent this problem, we have now designed 2-amino-2,3-dihydro-1*H*-cyclopenta-[*b*]anthracene-2-carboxylic acid (antAib), a new fluorescent α -amino acid residue which is based on a planar anthracene core and, like Bin, belongs to the class of $C_i^{\alpha} \rightarrow C_i^{\alpha}$ cyclized, $C^{\alpha,\alpha}$ -disubstituted glycines (effective β -turn and helix inducers in peptides⁴). The achiral antAib residue may be regarded either as a rigid analogue of the known 9-antAla⁵ (or its cyano derivative Flu^{1a}) and 2-antAla^{5b} residues,



Figure 1. Chemical structure of the antAib residue compared with Bin,² 9-antAla,⁵ and 2-antAla.^{5b}

Keywords: Anthracene; Aromatic amino acid; $C^{\alpha,\alpha}$ -Disubstituted glycine; Fluorescent amino acid.

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with the spatial disposition of the anthracene side-chain fluorophore relative to the α -carbon atom being completely defined, or as an anthracene-fused 1-aminocyclopentanecarboxylic acid (Ac₅c). Only quinonic derivatives of antAib have been synthesized previously by different methods.⁶

2. Results and discussion

For the synthesis of antAib, the known dimethyl anthracene-2.3-dicarboxylate 5 was used as a key intermediate. This compound was readily prepared by using an easily reproducible, published, procedure (Fig. 2),⁷ in which Friedel–Crafts acylation of 1,2,4-trimethylbenzene 1 with benzoyl chloride gave the resulting benzophenone derivative 2 in 67% yield. In the second step, the two-stage oxidation of all three methyl groups present in 2 afforded the tricarboxylic acid 3 (61%), which upon cyclization in concentrated sulfuric acid formed the anthraquinone dicarboxylic acid 4 (88%). Reduction of the quinone part of 4 with activated zinc dust and ammonium hydroxide gave 2,3-anthracene dicarboxylic acid as a soft solid precipitate which could not be collected easily by filtration (see Section 3). Therefore, it was not purified, but collected by centrifugation and then directly esterified in refluxing methanol (MeOH) containing 98% H₂SO₄ to afford the dimethyl ester 5 in 71% overall yield after extraction followed by crystallization from acetone/methanol.



Figure 2. Synthesis of dimethyl anthracene-2,3-dicarboxylate **5**. (i) C_6H_5COCl ; AlCl₃; 0 °C to rt; 67%. (ii) (1) 20% aq HNO₃; reflux; (2) 10% aq NaOH; KMnO₄; reflux; (3) H⁺; 62%. (iii) 98% H₂SO₄; 120–130 °C; 87%. (iv) (1) Activated Zn powder; 20% aq NH₄OH; reflux; (2) 98% H₂SO₄; MeOH; reflux (71%).

Reduction of the diester **5** was accomplished under various experimental conditions. The expected diol was not isolated, but treated directly with acetic anhydride (Ac₂O) in pyridine^{8,9} to afford the corresponding 2,3-(bis)-acetoxymethyl-anthracene **6** (Fig. 3). The use of lithium aluminum hydride in THF (tetrahydrofuran) as the reducing agent always resulted in partial over-reduction of the central ring with formation of 2,3-(bis)-acetoxymethyl-9,10-dihydro-anthracene **7** along with **6**, in a ratio depending on reaction time and temperature (Table 1). Large-scale separation of the desired compound **6** from the side product **7** by chromatography was not possible because of their close R_f values (as observed by analytical TLC). However, their separation by fractional crystallization from acetone allowed the recovery of pure **6** in 46% yield from the first crop.

To transform **5** into the corresponding diol without overreduction of the central ring, the best method we found was the use of sodium bis(2-methoxyethoxy)aluminum hydride (RedAl[®]) at low temperature, as proposed by Sun and Desper in a similar case.⁸ Here, another side product was formed, the mono-reduced 2-acetoxymethyl-3-methylanthracene **8**, but in a very minor proportion relative to **6** (3% vs 86% isolated yields) when the reaction was conducted at low temperature (-15 °C to +3 °C) for a short period of time (1 h 15 min) (Table 1). Furthermore, the separation of **8** from **6** by chromatography was easy. Subsequent crystallization from boiling MeOH/CH₂Cl₂ afforded analytically pure diacetate **6** in 81% yield.

Bromuration of the diacetate **6** with HBr in acetic acid⁹ readily furnished 2,3-(bis)-bromomethyl-anthracene **9** (Fig. 4) in 98% yield after crystallization from boiling MeOH/CH₂Cl₂. The dibromide **9** was then used as an electrophile for the bisalkylation of ethyl isocyanoacetate under the phase transfer conditions developed by Kotha and Brahmachary,^{6b} with potassium carbonate as a base and tetra-*n*-butylammonium (^{*n*}Bu₄N⁺) hydrogen sulfate as the catalyst, in acetonitrile at 80 °C. The resulting 2-isocyano-2,3-dihydro-1*H*-cyclopenta[*b*]anthracene-2-carboxylic acid ethyl ester **10a** was not isolated, but rather the crude reaction mixture was submitted to acid hydrolysis to afford directly the desired α -amino

Table 1. Product distribution (%) after reduction of the diester 5 followed by acetylation

Reduction conditions		6	7	8
LiAlH ₄ (THF) LiAlH ₄ (THF) LiAlH ₄ (THF) RedAl [®] (THF) RedAl [®] (THF) RedAl [®] (THF)	$\begin{array}{c} 0 \ ^{\circ}\text{C to rt} \ (2 \ h) \\ -14 \ ^{\circ}\text{C to} \ -2 \ ^{\circ}\text{C} \ (1 \ h) \\ -60 \ ^{\circ}\text{C to} \ -15 \ ^{\circ}\text{C} \ (2 \ h) \\ 0 \ ^{\circ}\text{C} \ (2 \ h \ 30 \ min) \\ 0 \ ^{\circ}\text{C} \ (1 \ h \ 15 \ min) \\ -15 \ ^{\circ}\text{C to} \ +3 \ ^{\circ}\text{C} \ (1 \ h \ 15 \ min) \end{array}$	$ \begin{array}{r} 30^{a} \\ 82^{a} 36^{b} \\ 85^{a} 46^{b} \\ 54^{b} \\ 73^{b} \\ 86^{b} \\ \end{array} $	70 ^a 18 ^a 15 ^a	10 ^b 5 ^b 3 ^b

⁴ Ratio determined by ¹H NMR.

Isolated yield.



Figure 3. Reduction of the diester 5 followed by acetylation. (i) LiAlH₄; THF; low temperature (see Table 1). (ii) RedAl[®] 3.5 M in toluene; THF; low temperature (see Table 1). (iii) Ac₂O; pyridine.



Figure 4. Synthesis of Boc–antAib–OH **13a** and Fmoc–antAib–OH **13b** from the diacetate **6**. (i) 33% HBr in AcOH; CH₂Cl₂; rt. (ii) CN–CH₂–COOEt (a series) or CN–CH₂–COO[']Bu (b series); K₂CO₃ or Cs₂CO₃; "Bu₄N⁺, HSO₄⁻ (cat); CH₃CN; 80 °C. (iii) 10 N aq HCl; abs EtOH; 0 °C to rt. (iv) Boc₂O; CH₃CN; rt (a series) or Fmoc–OSu; CH₃CN/CH₂Cl₂; rt (b series). (v) 1 N NaOH; MeOH/THF; rt (a series) or TFA/CH₂Cl₂ 1:2; 0 °C to rt (b series).

ester H-antAib-OEt 11a in 28% yield after chromatography. It may be pointed out that this relatively moderate yield, due to unidentified side reactions of the dibromide 9, is in agreement with the 29.7% yield10 previously obtained by Kotha et al.^{6b} in a similar synthesis of the parent anthraquinonefused (instead of anthracene-fused) amino ester. Treatment of 11a with di-tert-butyl dicarbonate in acetonitrile gave the fully protected derivative Boc-antAib-OEt (Boc, tertbutyloxycarbonyl; OEt, ethoxy) 12a in only 37% yield after chromatography. In later runs we preferred not to isolate the α -amino ester **11a** but to perform *N*-Boc-protection directly on the crude product obtained after acid hydrolysis, thus reducing purification steps, which resulted in a slight increase in the overall yield in 12a from 9 (18–25%). In a similar manner, the bis-alkylation of tert-butyl isocyanoacetate was conducted in acetonitrile at 80 °C, using cesium carbonate as a base and tetrabutylammonium hydrogen sulfate as the phase transfer catalyst, to afford 2-isocyano-2,3-dihydro-1*H*-cyclopenta[b]anthracene-2-carboxylic acid tertbutyl ester 10b in 29% yield after chromatography. Remarkably, treatment of 10b with a few drops of concentrated hydrochloric acid (ca. 10 N) in ethanol (EtOH)/dichloromethane, from 0 °C to room temperature for a few hours (monitored by TLC) allowed the selective acidolysis of the isonitrile function without cleavage of the tert-butyl ester, and furnished H-antAib-O'Bu (O'Bu, tert-butoxy) **11b** in 90% vield after chromatography. Protection of the amino function of 11b by the Fmoc (9-fluorenylmethyloxycarbonyl) group was performed using Fmoc-OSu (OSu, 1-oxysuccinimide) in dichloromethane at room temperature to afford Fmoc-antAib-O'Bu 12b in 86% yield. Finally, Cdeprotection of 12a by saponification of the ethyl ester function with 1 N NaOH in MeOH/THF gave Boc-antAib-OH 13a (89–96% yield), and C-deprotection of 12b by acidolysis of the tert-butyl ester function with TFA (trifluoroacetic

acid)/ CH_2Cl_2 gave **13b** (92% yield), both suitable for use in peptide synthesis.

Solution synthesis of di- and tripeptides based on antAib was carried out in order to investigate the coupling ability at both C- and N-termini of such a structurally constrained residue. Coupling of Boc-antAib-OH and HCl·L-H-Ala-OMe was performed by EDC [N-ethyl, N'-(3-dimethylaminopropyl)-carbodiimide]/HOAt (7-aza-1-hydroxy-1,2, 3-benzotriazole)¹¹ C-activation to furnish Boc-antAib-L-Ala-OMe 14a (Fig. 5) in 50-60% yield. This dipeptide was N^{α} -deprotected with TFA/CH₂Cl₂ 1:3 and the resulting TFA·H-antAib-L-Ala-OMe (not isolated) was coupled with the urethane-protected N-carboxyanhydride (UNCA)¹² Fmoc-L-Ala-NCA, to afford the tripeptide Fmoc-L-AlaantAib-L-Ala-OMe 15a in 83% overall yield. These two methods are known to be efficient in difficult cases involving sterically demanding C^{α} -tetrasubstituted α -amino acids.¹³ It is also interesting to note that, as previously observed in a similar case,¹⁴ coupling of N^{α} -deprotected **14a** with the more hindered Boc–Aib–NCA (Aib, α -aminoisobutyric acid) afforded the tripeptide Boc-Aib-antAib-L-Ala-OMe 16a in only 14% overall yield, because of a competitive cyclization to the 2,5-dioxopiperazine cyclo-[antAib-L-Ala] 17a, isolated in 48% yield. One can expect therefore that in general cases where antAib is at the N-terminal position of a peptide, but not of a dipeptide methyl ester (where a fast cyclization reaction is favored), coupling at its amino function with an additional C^{α} -tetrasubstituted α -amino acid, should occur efficiently.

We have recorded the UV absorption spectra of Boc–antAib–OEt **12a** and Boc–antAib–OH **13a** and their fluorescence spectral signatures in ethanol solution. The symmetry-allowed, intense, $S_0 \rightarrow S_1$ transition of the



Figure 5. Solution synthesis of di- and tripeptides based on antAib. (i) HCl·H–L-Ala–OMe; NMM (*N*-methylmorpholine); EDC; HOAt; THF/CH₂Cl₂; rt. (ii) TFA/CH₂Cl₂ (1:3); 0° C. (iii) Fmoc–L-Ala–NCA; DIEA (*N*,*N*,*N*-diisopropylethylamine); THF; 0° C to rt. (iv) Boc–Aib–NCA; DIEA; THF; 60° C.

anthracene chromophore is evident as a Frank–Condon vibronic progression with an origin (0,0) at 378 nm and additional peaks (0,1–0,4) separated by ca. 16 nm (λ_{max} = 359 nm)¹⁵ (Fig. 6). The emission spectra (λ_{exc} =359 nm) have an origin at 384–385 nm, a maximum intensity at 407–408 nm, and two other peaks of the vibronic progression clearly observed at longer wavelengths (Fig. 7).

In conclusion, the syntheses of the protected derivatives Boc–antAib–OEt and Fmoc–antAib–O'Bu were achieved in seven steps from 1,2,4-trimethylbenzene. Saponification of the ester function of Boc–antAib–OEt and acidolysis of Fmoc–antAib–O'Bu afforded the corresponding *N*-protected amino acids, suitable for peptide elongation using either Boc or Fmoc strategies. Solution peptide syntheses of the tripeptides Fmoc–L-Ala–antAib–L-Ala–OMe and Boc–Aib– antAib–L-Ala–OMe demonstrated the coupling efficiency at both C- and N-termini of the structurally constrained antAib residue. The fluorescence spectra of Boc–antAib– OEt and Boc–antAib–OH suggest that the antAib residue may represent a novel useful spectroscopic probe in studies of peptide molecules by virtue of the appearance of its bands



Figure 6. UV absorption spectrum (>300 nm region) of Boc–antAib–OEt **12a** (solid line) and Boc–antAib–OH **13a** (dotted line) in ethanol solution. Amino acid derivative concentration: 1×10^{-4} M.



Figure 7. Fluorescence spectra in the 370–500 nm range of Boc–antAib– OEt **12a** (solid line) and Boc–antAib–OH **13a** (dotted line) in ethanol solution: λ_{exc} =359 nm. Amino acid derivative concentration: 1×10^{-6} M.

at much longer wavelengths than those typical of coded aromatic α -amino acids. To determine antAib preferred conformation, peptides based on this residue in combination with L-Ala to the hexamer level are currently being synthesized in our groups. Secondary structures involving β -turns/3₁₀-helices are expected to be efficiently induced by antAib as it is a member of the family of $C_i^{\alpha} \rightarrow C_i^{\alpha}$ cyclized, $C^{\alpha,\alpha}$ -disubstituted glycines,⁴ thus allowing its exploitation as a fluorophore in photophysical studies of rigid, folded, peptide architectures.

Altogether, we believe that the antAib residue will expand the scope of fluorescence analysis of peptide conformations and interactions in solution, as it represents a *unique label with frozen main-chain and side-chain rotational freedoms*, in contrast with all fluorescent probes proposed so far. That may be of main interest for the design of conformationally constrained bioactive peptide systems.¹⁶

3. Experimental

Melting points were measured on a Mettler apparatus with a final temperature raise of 3 °C/min or by means of a capillary tube immersed in an oil bath (Tottoli apparatus, Büchi) and are uncorrected. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker WM300 spectrometer operating at 300 MHz and 77 MHz, respectively, the solvent being used as the internal standard: CDCl₃ (¹H: δ =7.27 ppm; ¹³C: δ =77.00 ppm), CD₃COCD₃ (¹H: δ =2.05 ppm; ¹³C: $\delta = 29.80 \text{ ppm}$, CD₃SOCD₃ (¹H: $\delta = 2.50 \text{ ppm}$; ¹³C: $\delta =$ 39.50 ppm). Splitting patterns are abbreviated as follows: (s) singlet, (d) doublet, (t) triplet, (q) quartet, (m) multiplet. Elemental analyses were performed by the C.N.R.S. Service of Microanalyses in Gif-sur-Yvette (France). Mass spectra (electrospray mode) were recorded by Vincent Steinmetz (Institut Lavoisier), and high-resolution mass spectra by Nicole Morin (Service of mass spectrometry, ENS, Paris). Analytical TLC and preparative column chromatography were performed on Kieselgel F 254 and Kieselgel 60 (0.040-0.063 mm) (Merck), respectively, with the following eluant systems: CH₂Cl₂ (I); 2.5% MeOH-97.5% CH₂Cl₂ (II); 2.5% EtOAc-97.5% CH₂Cl₂ (III); 5% EtOAc-95% CH₂Cl₂ (IV); 20% EtOAc-80% CH₂Cl₂ (V); 50% EtOAc-50% CH₂Cl₂ (VI); 5% MeOH-95% CH₂Cl₂ (VII); 10% MeOH-90% CH₂Cl₂ (VIII). UV light $(\lambda = 254 \text{ nm})$ allowed visualization of the spots after TLC runs for all compounds. Except when noted, all starting materials and solvents were obtained from commercial suppliers and were used as received. Fmoc-L-Ala-NCA and Boc-Aib-NCA were purchased from Fluka and Isochem, respectively.

3.1. 2,4,5-Trimethylbenzophenone (2)

A mixture of 1,2,4-trimethylbenzene **1** (20 g; 166.6 mmol) and AlCl₃ (23.4 g; 175 mmol) in CH₂Cl₂ (20 mL) was treated with benzoyl chloride (23.4 g; 166.5 mmol) as described in the literature,⁷ to yield 24.9 g (67%) of pure **2** as a colorless liquid after vacuum distillation. Bp 152–155 °C/ca. 0.3 Torr (lit.⁷ bp 130 °C/0.15 Torr). ¹H NMR (CDCl₃): δ 7.81 [m, 2H, ArH], 7.58 [m, 1H, ArH], 7.46 [m, 2H, ArH], 7.12 [s, 1H, ArH], 7.07 [s, 1H, ArH], 2.30 [s, 3H, ArCH₃], 2.29 [s, 3H, ArCH₃], 2.25 [s, 3H, ArCH₃].

¹³C NMR (CDCl₃): δ 198.2 [C=O], 138.9, 138.0, 135.8, 134.2, 133.0, 132.5, 132.2, 129.9, 129.7, 129.1, 128.5, 128.1 [C_{Ar}], 19.35, 19.29, 18.8 [ArCH₃].

3.2. Benzophenone-2,4,5-tricarboxylic acid (3)

A mixture of 2,4,5-trimethylbenzophenone **2** (19.7 g; 88 mmol) and 20% HNO₃ (150 mL) was refluxed with stirring for 5 days. The resulting thick semisolid was decanted, rinsed with cold water (2×75 mL), dissolved in boiling 10% NaOH (200 mL), and treated with KMnO₄ (55.3 g; 350 mmol) as described in the literature,⁷ to yield 17.0 g (61%) of pure **3** obtained as white crystals. Mp 224 °C (lit.⁷ mp 281–283 °C). ¹H NMR (CD₃COCD₃): δ 8.47 [s, 1H, ArH], 7.76 [m, 2H, ArH], 7.75 [s, 1H, ArH], 7.67–7.46 [m, 3H, ArH]. ¹³C NMR (CD₃COCD₃): δ 195.7 [C=O], 167.9, 167.4, 165.8 [COOH]; 145.5, 137.7, 137.6, 134.1, 133.8, 132.2, 131.7, 130.0, 129.4, 128.6 [C_{Ar}].

3.3. Anthraquinone-2,3-dicarboxylic acid (4)

The triacid **3** (4.20 g; 13.4 mmol) was treated with 98% H_2SO_4 (42 g) at 120–130 °C for 3 h as described in the literature,⁷ to yield 3.49 g (88%) of pure **4** as a pale brown solid. Mp >300 °C (lit.⁷ mp >310 °C). ¹H NMR (CD₃SOCD₃): δ 8.36 [s, 2H, ArH], 8.26–8.19 [m, 2H, ArH], 8.00–7.94 [m, 2H, ArH]. ¹³C NMR (CD₃SOCD₃): δ 181.4 [C=O], 167.6 [COOH]; 137.5, 134.9, 134.2, 132.9, 127.0, 126.9 [C_{Ar}].

3.4. Dimethyl anthracene-2,3-dicarboxylate (5)

Following the literature procedure,⁷ the diacid 4 (2.00 g; 6.76 mmol) was added by portions to a solution of 20% NH₄OH (100 mL) with stirring at room temperature. A clear red solution was obtained after ca. 15 min, to which was added by portions activated zinc dust (7.50 g). The resulting blood-red mixture was refluxed until the color was gone (ca. 2 h) and then filtered while hot. The solid residue was refluxed with 20% NH₄OH (50 mL) for 2 h and filtered while hot. The combined filtrates were cooled on an ice bath and then acidified to pH <1 with 10 N HCl, which resulted in the precipitation of a yellow colloidal solid which slowly settled overnight at room temperature. Attempted filtration on a Büchner was very slow and difficult. Therefore, the solid was collected by centrifugation, and then repeatedly dried by evaporation in vacuo at 50 °C after addition of methanol. The so-obtained crude anthracene-2,3-dicarboxylic acid (2.14 g) was pure by ¹H NMR (CD₃SOCD₃): δ 8.77 [s, 2H, ArH], 8.46 [s, 2H, ArH], 8.17-8.12 [m, 2H, ArH], 7.64–7.59 [m, 2H, ArH] with only the presence of NH₄Cl $(\delta 7.18 \text{ [t, } J=51.0 \text{ Hz]})$ as contaminant. Ten similar runs starting with in total 19.25 g (65.0 mmol) of diacid 4 gave 21.80 g of such a mixture of anthracene-2,3-dicarboxylic acid and NH₄Cl. To this mixture (10.10 g) was added MeOH (400 mL) and 98% H₂SO₄ (10 mL). The resulting yellow suspension was stirred and refluxed. After ca. 1 h, a clear orange-brown solution was obtained, which was refluxed for 5 days to give a suspension again. The mixture was cooled to 0 °C, the solid was filtered, abundantly washed with MeOH, air dried (weight 6.27 g), and then dissolved in a mixture of H₂O (400 mL) and CH₂Cl₂ (600 mL) with stirring. The decanted CH₂Cl₂ solution was washed with 5% NaHCO₃ (100 mL), H₂O (2×400 mL), dried (MgSO₄), filtered, and evaporated in vacuo to afford 5.76 g of crude diester 5 as a solid, which was pure by ¹H NMR and TLC. The original mother liquor (MeOH solution) was concentrated in vacuo at 40 °C to ca. 150 mL, H₂O (700 mL) added, and the mixture was extracted with CH₂Cl₂ (250 mL). The decanted CH₂Cl₂ solution was washed with 5% NaHCO₃ (100 mL) and H₂O (2×400 mL) as above, dried (MgSO₄), filtered, and evaporated in vacuo to afford 1.01 g of impure diester 5. A second similar run starting from the remaining mixture of diacid 4 and NH₄Cl (11.70 g) gave 7.39 g of pure diester 5 after extraction of the filtered precipitate and 1.39 g of impure compound from the mother liquor. The combined pure samples of diester (13.14 g) were dissolved in boiling acetone (400 mL) and the solution concentrated to ca. 60 mL. Crystallization occurred from the boiling solution to which methanol (ca. 60 mL) was added by portions in order to increase the quantity of crystals. The mixture was concentrated again to ca. 60 mL, and then cooled to room temperature. The crystals were filtered, abundantly washed with methanol, and air dried (weight 11.96 g). More crystals (1.60 g) were obtained from the mother liquor and by repeated crystallization of the combined impure samples, to give a total of 13.56 g (71%) of analytically pure diester 5, obtained as yellow-orange crystals. Mp 152 °C (lit.⁷ mp 149–151 °C). R_f 0.18 (I). ¹H NMR (CDCl₃): δ 8.52 [s, 2H, ArH], 8.45 [s, 2H, ArH], 8.09–8.04 [m, 2H, ArH], 7.60–7.55 [m, 2H, ArH]. ¹³C NMR (CDCl₃): δ 168.0 [C=O], 132.9, 131.1, 130.2, 128.2, 127.8, 127.4, 126.7 [C_{Ar}], 52.5 [OCH₃].

3.5. Reduction of dimethyl anthracene-2,3-dicarboxylate

(a) With lithium aluminum hydride: To a suspension of LiAlH₄ (0.190 g; 5 mmol) in THF (15 mL) magnetically stirred at 0 °C was added the diester 5 (0.294 g; 1 mmol) by portions under an argon atmosphere. The reaction mixture was stirred at room temperature for 2 h, cooled to 0 °C, and quenched by dropwise addition of saturated aqueous Na₂SO₄ (ca. 2 mL). After stirring at room temperature for 15 min, the mixture was filtered on glass wool and the filtrate was evaporated to dryness in vacuo at 40 °C. The residue was repeatedly evaporated again in vacuo after addition of methanol in order to remove water. The obtained crude product was dissolved in pyridine (10 mL) and acetic anhydride (1 mL) was added. The resulting solution was stirred at room temperature overnight and then evaporated to dryness in vacuo at 40 °C. The residue was taken up in ethyl acetate (100 mL) and 0.5 N aq HCl (100 mL) with stirring. The decanted organic phase was washed with 0.5 N aq HCl (100 mL), then with H_2O (2×100 mL), dried (MgSO₄), filtered, and evaporated in vacuo. The crude product (0.175 g) presented a single spot on TLC with either eluant (I), (II), (III) or (IV), but contained compounds 6 and 7 in the ratio 30:70, determined by integration of their respective ArCH₂O and OCOCH₃ singlets in ¹H NMR (vide infra). Preparative TLC on silica gel with eluant (III) gave 0.105 g of the same mixture 6 (30%) and 7 (70%). Fractional crystallization from a concentrated acetone solution (ca. 1 mL) gave 0.012 g (4%) of pure **6** as pale yellow crystals.

In another run, the diester 5(0.294 g; 1 mmol) was treated by LiAlH₄ (0.190 g; 5 mmol) in THF (15 mL) in the same experimental conditions and workup as above, except that

the reaction was carried out at -14 °C to -2 °C for 1 h. The crude product obtained after acetylation (0.213 g) contained **6** and **7** in the ratio 82:18 by ¹H NMR. Fractional crystallization of this crude product from concentrated acetone (ca. 1 mL) gave 0.116 g (36%) of pure **6** as pale yellow crystals.

In another run, the diester **5** (1.470 g; 5 mmol) was treated by LiAlH₄ (0.950 g; 25 mmol) in THF (75 mL) in the same experimental conditions and workup as above, except that the reaction was carried out at -60 °C to -15 °C for 2 h. The crude product obtained after acetylation (1.486 g) contained compounds **6** and **7** in the ratio 85:15 by ¹H NMR. Fractional crystallization of this crude product from concentrated acetone (ca. 5 mL) gave 0.746 g (46%) of pure **6** as pale yellow crystals.

(b) With sodium bis(2-methoxyethoxy)aluminum hydride: To a solution of diester 5 (2.940 g; 10 mmol) in THF (100 mL) cooled to 0 °C and stirred under an argon atmosphere was added dropwise by syringe 16 mL (56 mmol) of ca. 3.5 M solution of RedAl[®] in toluene (Acros) over a period of 30 min. The resulting red-brown solution was stirred at 0 °C for 2 h, and then quenched by dropwise addition of aq 5% H₂SO₄ (100 mL). The mixture was poured into a separating funnel containing 5% H₂SO₄ (200 mL) and EtOAc (500 mL). After shaking and decantation, a large amount of foam at the interface between the milky yellow organic phase and the clear colorless aqueous acidic phase was present, due to the low solubility of the expected 2,3-bis(hydroxymethyl)-anthracene. The separated organic phase and the foam at the interface were washed with H₂O (500 mL), decanted, and then directly (without addition of drving agent) evaporated to dryness in vacuo. The residue was repeatedly evaporated again in vacuo after addition of methanol in order to remove water. The obtained crude product (2.454 g) was dissolved in pyridine (100 mL) and acetic anhydride (20 mL) was added. The resulting clear orange solution was magnetically stirred at room temperature for 20 h and then evaporated to dryness in vacuo at 40 °C. The residue was taken up in ethyl acetate (400 mL) and 0.5 N aq HCl (300 mL) with stirring. The decanted organic phase was washed with 0.5 N aq HCl (100 mL), then with brine (2×400 mL), dried (MgSO₄), filtered, and evaporated in vacuo. The crude product (3.161 g) was chromatographed on a 2.3×54 cm column of silica gel with eluant (IV) to afford the desired pure 2,3-bis(acetoxymethyl)-anthracene 6 (1.752 g; 54%) and pure 2-acetoxymethyl-3-methylanthracene 8 (0.272 g; 10%) as a side product.

In another run, the diester **5** (2.940 g; 5 mmol) in THF (100 mL) was treated by a 3.5 M solution of RedAl[®] in toluene (16 mL; 56 mmol) in the same experimental conditions and workup as above, except that the reaction was conducted at 0 °C for 15 min (addition) and then for a further 1 h. Column chromatography of the crude product obtained after acetylation (3.198 g) gave **6** (2.354 g; 73%) and **8** (0.140 g; 5%).

In another run, the diester **5** (3.633 g; 12.4 mmol) in THF (125 mL) was treated by a 3.5 M solution of RedAl[®] in toluene (19 mL; 66 mmol) in the same experimental conditions and workup as above, except that the reaction was conducted at -15 °C to -13 °C for 15 min (addition) and then

at -13 °C to +3 °C for a further 1 h. Column chromatography of the crude product obtained after acetylation (3.917 g) gave **6** (3.437 g; 86%) and **8** (0.114 g; 3%). The pure compound **6** (3.437 g) was dissolved in boiling CH₂Cl₂ (100 mL) (water bath at 80 °C) and MeOH (50 mL) was added. The solution was concentrated to ca. 50 mL upon boiling (crystallization started to occur from the boiling solution). Methanol (50 mL) was added, the mixture was concentrated again to ca. 30 mL upon boiling, and then cooled to room temperature. The resulting crystals were filtered, abundantly washed with MeOH, and air dried to give 3.203 g (81%) of analytically pure **6**. Similar crystallization of the side product **8** (0.553 g; combined samples from several runs) from boiling MeOH (CH₂Cl₂) gave 0.384 g of yellow crystals.

3.6. 2,3-Bis(acetoxymethyl)-anthracene (6)

Yellow crystals. Mp 185 °C. R_f 0.2–0.3 (I), 0.75 (II), 0.35 (III), 0.50 (IV). ¹H NMR (CDCl₃): δ 8.43 [s, 2H, ArH], 8.04 [s, 2H, ArH], 8.04–8.00 [m, 2H, ArH], 7.53–7.48 [m, 2H, ArH], 5.39 [s, 4H, ArCH₂O], 2.17 [s, 6H, OCOCH₃]. ¹³C NMR (CDCl₃): δ 170.6 [C=O], 132.2, 131.3, 130.8, 129.7, 128.2, 126.4, 125.8 [C_{Ar}], 64.6 [ArCH₂O], 20.9 [OCOCH₃]. Anal. Calcd for C₂₀H₁₈O₄ (322.344): C, 74.52; H, 5.63. Found: C, 74.92; H, 5.65.

3.7. 2,3-Bis(acetoxymethyl)-9,10-dihydroanthracene (7)

Not isolated. Characterized by ¹H and ¹³C NMR in mixtures with **6**. R_f 0.2–0.3 (I), 0.75 (II), 0.35 (III), 0.50 (IV). ¹H NMR (CDCl₃): δ 7.36 [s, 2H, ArH], 7.32–7.27 [m, 2H, ArH], 7.25–7.20 [m, 2H, ArH], 5.22 [s, 4H, ArCH₂O], 3.95 [s, 4H, ArCH₂Ar], 2.11 [s, 6H, OCOCH₃]. ¹³C NMR (CDCl₃): δ 170.6 [C=O], 137.4–125.7 [C_{Ar}], 63.7 [ArCH₂O], 35.6 [ArCH₂Ar], 20.9 [OCOCH₃].

3.8. 2-Acetoxymethyl-3-methyl-anthracene (8)

Yellow crystals. Mp 194 °C. R_f 0.70 (IV). ¹H NMR (CDCl₃): δ 8.43 [s, 1H, ArH], 8.40 [s, 1H, ArH], 8.02–7.97 [m, 2H, ArH], 7.98 [s, 1H, ArH], 7.80 [s, 1H, ArH], 7.49–7.44 [m, 2H, ArH], 5.32 [s, 2H, ArCH₂O], 2.55 [s, 3H, ArCH₃], 2.18 [s, 3H, OCOCH₃]. ¹³C NMR (CDCl₃): δ 170.9 [C=O], 133.9, 132.5, 132.1, 131.7, 131.5, 130.3, 128.5, 128.2, 128.1, 126.2, 125.5, 125.1, 125.0 [C_{Ar}], 65.2 [ArCH₂O], 21.0 [OCOCH₃], 19.5 [ArCH₃].

3.9. 2,3-Bis(bromomethyl)-anthracene (9)

To a solution of diacetate **6** (3.222 g; 10 mmol) in CH₂Cl₂ (200 mL) was added a 33 wt % solution of hydrogen bromide in acetic acid (Aldrich) (20 mL; ca. 110 mmol). The solution, which became more and more turbid, was stirred at room temperature for 24 h. Water (250 mL) was added, the reaction mixture was transferred to a separating funnel, and extracted with CH₂Cl₂ (800 mL necessary for complete solubilization). The separated CH₂Cl₂ solution was washed with 5% NaHCO₃ (150 mL) and then H₂O (2×150 mL), dried (MgSO₄), filtered, and evaporated in vacuo. The obtained crystalline crude product (3.680 g) was stirred in boiling CH₂Cl₂ (200 mL) but could not be totally solubilized. Methanol (50 mL) was added and the mixture was concentrated to ca. 30 mL with almost complete crystallization occurring from the boiling solution. After cooling to room temperature, the crystals were filtered, abundantly washed with MeOH, and air dried to afford 3.573 g (98%) of analytically pure dibromide **9** as yellow crystals. Mp 225 °C. R_f 0.85 (III). ¹H NMR (CDCl₃): δ 8.39 [s, 2H, ArH], 8.04 [s, 2H, ArH], 8.03–7.99 [m, 2H, ArH], 7.53–7.49 [m, 2H, ArH], 4.95 [s, 4H, ArCH₂Br]. ¹³C NMR (CDCl₃): δ 133.1, 132.4, 131.2, 131.0, 128.2, 126.6, 126.1 [C_{Ar}], 31.6 [ArCH₂Br]. HRMS (DCI⁺). Calcd [M+H]⁺ for C₁₆H₁₃ ⁷⁹Br₂: 362.9384. Found: 362.9388. Calcd [M+H]⁺ for C₁₆H₁₃ ⁷⁹Br⁸¹Br: 364.9364. Found: 364.9373. Calcd [M+H]⁺ for C₁₆H₁₃ ⁸¹Br₂: 366.9346. Found: 366.9350. Anal. Calcd for C₁₆H₁₂Br₂ (364.088): C, 52.78; H, 3.32. Found: C, 52.93; H, 3.29.

3.10. 2-Isocyano-2,3-dihydro-1*H*-cyclopenta[*b*]anthracene-2-carboxylic acid *tert*-butyl ester (C=antAib-O^tBu 10b)

To a suspension of 9 (1.00 g; 2.75 mmol), tetrabutylammonium hydrogen sulfate (0.42 g; 1.24 mmol) and Cs₂CO₃ (5.38 g; 16.5 mmol) in CH₃CN (150 mL) was added tertbutyl isocyanoacetate (0.60 mL, 4.12 mmol). The mixture was stirred under argon at 75-80 °C for 48 h and then filtered through sintered glass. The filtrate was evaporated in vacuo and the residue dissolved in CH₂Cl₂ (150 mL). The organic layer was washed with water, dried over magnesium sulfate, and evaporated to dryness. The crude product was purified by column chromatography on silica gel using CH₂Cl₂ as eluant to give 0.275 g (29%) of pure **10b** as a pale yellow solid. Crystallization of an aliquot from cyclohexane/CH₂Cl₂ afforded analytically pure crystals (needles). Mp 203 °C. R_f 0.55 (I). ¹H NMR (CDCl₃): δ 8.37 [s, 2H, ArH], 8.00–7.97 [m, 2H, ArH], 7.85 [s, 2H, ArH], 7.48–7.45 [m, 2H, ArH], 3.81 and 3.61 [d, J=16.4 Hz, 2H and d, J=16.4 Hz, 2H, ArCH₂], 1.54 [s, 9H, CH₃ O'Bu]. ¹³C NMR (CDCl₃): δ 167.3 [C=O], 137.1, 131.9, 131.7, 128.3, 126.2, 125.6, 123.3 [C_{Ar}], 111.9 [C=N-], 84.3 [O-C O^tBu], 77.4 [C^{α}], 45.5 [ArCH₂], 28.0 [CH₃ O'Bu]. Anal. Calcd for C₂₃H₂₁NO₂ (343.406): C, 80.44; H, 6.16; N, 4.08. Found: C, 80.43; H, 6.13; N, 3.88.

3.11. 2-Amino-2,3-dihydro-1*H*-cyclopenta[*b*]anthracene-2-carboxylic acid ethyl ester (H–antAib–OEt 11a)

To a suspension of 9 (0.182 g; 0.5 mmol), tetrabutylammonium hydrogen sulfate (0.068 g; 0.2 mmol) and K₂CO₃ (1.38 g; 10 mmol) in CH₃CN (25 mL) was added ethyl isocyanoacetate (0.55 mL; 5 mmol). The mixture was stirred under argon at 75-80 °C for 20 h, then cooled to room temperature, and filtered through sintered glass. The solid was abundantly washed with CH₂Cl₂ and the filtrate was evaporated in vacuo. To the residue (containing C=antAib-OEt 10a, not isolated) was added CH₂Cl₂ (10 mL), abs EtOH (25 mL) and 10 N HCl (1 mL). The mixture was magnetically stirred at room temperature for 2 h and then diluted with CH₂Cl₂ (100 mL). Water (150 mL) was added, and the mixture was made alkaline by slow addition of NaHCO₃ with stirring. The separated CH₂Cl₂ solution was washed with H₂O (2×100 mL), dried (MgSO₄), filtered, and evaporated in vacuo. The crude product was purified on a preparative TLC plate of silica gel with eluant (VI) to afford 0.043 g (28%) of α -amino ester **11a**. R_f 0.05 (IV); 0.20 (V). ¹H NMR (CDCl₃): δ 8.33 [s, 2H, ArH], 8.00– 7.95 [m, 2H, ArH], 7.82 [s, 2H, ArH], 7.46–7.41 [m, 2H, ArH], 4.27 [q, *J*=7.1 Hz, 2H, CH₂ OEt], 3.71 and 3.08 [d, *J*=16.3 Hz, 2H, ArCH₂ and d, *J*=16.3 Hz, 2H, ArCH₂], 2.02 [br s, 2H, NH₂], 1.33 [t, *J*=7.1 Hz, 3H, CH₃ OEt]. ¹³C NMR (CDCl₃): δ 172.5 [C=0], 139.8, 131.7, 131.4, 128.0, 125.5, 125.0, 122.8 [C_{Ar}], 65.4 [C^{α}], 61.5 [CH₂ OEt], 45.4 [ArCH₂], 14.2 [CH₃ OEt]. HRMS (FAB⁺) of **11a**·CF₃CO₂H. Calcd [M+H]⁺ for C₂₀H₂₀NO₂: 306.1494. Found: 306.1487.

3.12. 2-Amino-2,3-dihydro-1*H*-cyclopenta[*b*]anthracene-2-carboxylic acid *tert*-butyl ester (H–antAib–O'Bu 11b)

The isonitrile 10b (0.103 g; 0.300 mmol) was dissolved in CH₂Cl₂ (20 mL) and ethanol (20 mL) was added. The solution was cooled to 0 °C and 10 N HCl (0.5 mL) was added. The reaction mixture was allowed to warm to room temperature and stirred for ca. 2 h, until total hydrolysis had occurred (TLC monitoring). Water (150 mL) and CH₂Cl₂ (150 mL) were added, and the mixture was made alkaline by slow addition of a large excess of NaHCO₃ (0.840 g; 10 mmol) with stirring. The separated CH₂Cl₂ solution was washed with water (2×100 mL), dried (MgSO₄), filtered, and evaporated in vacuo. The crude product was chromatographed on a column of silica gel with eluant (V) to afford 0.090 g (90%) of pure 11b as a pale yellow solid. Mp 227 °C. R_f 0.31 (V). ¹H NMR (CDCl₃): δ 8.33 [s, 2H, ArH], 8.00-7.96 [m, 2H, ArH], 7.81 [s, 2H, ArH], 7.47-7.42 [m, 2H, ArH], 3.66 and 3.05 [d, J=16.3 Hz, 2H, ArCH₂ and d, J=16.3 Hz, 2H, ArCH₂], 1.86 [br s, 2H, NH₂], 1.51 [s, 9H, CH₃ O'Bu]. ¹³C NMR (CDCl₃): δ 175.7 [C=O], 140.4, 131.9, 131.5, 128.2, 125.7, 125.1, 122.9 $[C_{Ar}]$, 81.6 [O–C O^{*t*}Bu], 66.1 [C^{α}], 45.6 [ArCH₂], 28.2 [CH₃ O^tBu]. Anal. Calcd for C₂₂H₂₃NO₂ (333.412): C, 79.25; H, 6.95; N, 4.20. Found: C, 79.07; H, 6.98; N, 4.05.

3.13. 2-*tert*-Butyloxycarbonylamino-2,3-dihydro-1*H*-cyclopenta[*b*]anthracene-2-carboxylic acid ethyl ester (Boc–antAib–OEt 12a)

(a) A solution of the α -amino ester **11a** (0.034 g; 0.11 mmol) and Boc₂O (0.050 g; 0.22 mmol) in CH₃CN (2 mL) was stirred at room temperature for 3 days and then evaporated in vacuo. The crude product was chromatographed on a preparative TLC plate of silica gel with eluant (III) to afford 0.0167 g (37%) of pure **12a**.

(b) In other experiments on a larger scale, without isolation/ characterization of the isonitrile and α -amino ester intermediates, a mixture of dibromide **9** (0.364 g; 1 mmol), tetrabutylammonium hydrogen sulfate (0.136 g; 0.4 mmol), K₂CO₃ (2.76 g; 10 mmol), and ethyl isocyanoacetate (0.30 mL; 2.75 mmol) in CH₃CN (50 mL) was reacted in the same experimental conditions and workup as above (in Section 3.10). The crude product was submitted to direct acidic hydrolysis with 10 N HCl (1 mL) in CH₂Cl₂ (10 mL) and abs EtOH (25 mL), also as above. The crude product obtained after extraction (containing the α -amino ester **11a**) was reacted with Boc₂O (0.347 g; 1.59 mmol) in CH₃CN (5 mL) and CH₂Cl₂

(10 mL) at room temperature for 5 days and then evaporated in vacuo. The crude product was chromatographed on a 2.3×27 cm column of silica gel with eluant (III), to afford 0.102 g (25%) of pure N-Boc-protected α -amino ester **12a**. In another run on a larger scale, a mixture of dibromide 9 (1.820 g; 5 mmol), tetrabutylammonium hydrogen sulfate (0.678 g; 2 mmol), K₂CO₃(13.8 g; 100 mmol), and ethyl isocyanoacetate (1.5 mL; 2.75 mmol) in CH₃CN (250 mL) was reacted in the same experimental conditions and workup as above. The crude product was submitted to direct acidic hydrolvsis with 10 N HCl (10 mL) in CH₂Cl₂ (75 mL) and abs EtOH (175 mL), also as above. The crude product obtained after extraction was reacted with Boc₂O (1.816 g; 8.33 mmol) in CH₃CN (25 mL) and CH₂Cl₂ (50 mL) at room temperature for 4 days, then diluted with CH₂Cl₂ (ca. 200 mL), filtered through Celite, and evaporated in vacuo. The crude product was dissolved in CH₂Cl₂ (75 mL) and chromatographed on a 3×42 cm column of silica gel with eluant (III) to afford 0.373 g (18%) of pure 12a. Several other runs in the same experimental conditions and workup gave a similar yield. The pure compound 12a from several runs (0.514 g) was dissolved in CH₂Cl₂ (25 mL) and EtOAc (5 mL) was added. The clear yellow solution was concentrated under a slight vacuum at 60 °C to ca. 4 mL where crystallization started to occur. Methanol (10 mL) was added and the mixture was concentrated again to ca. 2 mL, then cooled at +4 °C for 3 h. More MeOH (10 mL) was added, the crystals were filtered, abundantly washed with MeOH, and air dried to give 0.418 g (81%) of analytically pure 12a as a yellow solid. Another similar crystallization process starting from pure **12a** (0.722 g) gave 0.598 g of analytically pure crystals. Mp 221 °C. R_f 0.30 (III); 0.90 (V). ¹H NMR (CDCl₃): δ 8.33 [s, 2H, ArH], 7.99–7.96 [m, 2H, ArH], 7.79 [s, 2H, ArH], 7.47-7.43 [m, 2H, ArH], 5.11 [br s, 1H, NH], 4.27 [q, J=7.1 Hz, 2H, CH₂ OEt], 3.77 and 3.42 [d, J=16.9 Hz, 2H, ArCH₂ and br d, J=16.7 Hz, 2H, ArCH₂], 1.45 [s, 9H, CH₃ Boc], 1.29 [t, J=7.1 Hz, 3H, CH₃ OEt]. ¹³C NMR (CDCl₃): δ 173.2 [C=O], 155.0 [C=O Boc], 139.0, 131.6, 131.4, 128.0, 125.6, 125.1, 122.6 [C_{Ar}], 80.1 [C-O Boc], 66.4 [C^a], 61.6 [CH₂ OEt], 43.1 [ArCH₂], 28.2 [CH₃ Boc], 14.1 [CH₃ OEt]. ESI⁺ MS *m*/*z* (relative intensity): 428.3 (100) [M+Na]⁺, 833.5 (58) [2M+Na]⁺. HRMS (FAB⁺). Calcd [M]⁺ for C₂₅H₂₇NO₄: 405.1940. Found: 405.1953. Calcd $[M+H]^+$ for $C_{25}H_{28}NO_4$: 406.2018. Found: 406.2014. Anal. Calcd for C₂₅H₂₇NO₄ (405.474): C, 74.05; H, 6.71; N, 3.45. Found: C, 73.87; H, 6.95; N, 3.71.

3.14. 2-(9-Fluorenylmethoxycarbonylamino)-2,3-dihydro-1*H*-cyclopenta[*b*]anthracene-2-carboxylic acid *tert*-butyl ester (Fmoc–antAib–O^tBu 12b)

(a) A solution of the α -amino ester **11b** (0.120 g; 0.36 mmol) and Fmoc–OSu (0.146 g; 0.43 mmol) in CH₂Cl₂ (10 mL) was magnetically stirred at room temperature for 48 h, and then diluted with CH₂Cl₂ (100 mL). The CH₂Cl₂ solution was washed with 0.5 N HCl (100 mL), 5% NaHCO₃ (100 mL) and then H₂O (2×100 mL), dried (MgSO₄), filtered, and evaporated in vacuo. The crude product was chromatographed on a column of silica gel with eluant (I) to afford 0.172 g (86%) of pure **12b** as a pale yellow solid.

(b) In other experiments on a larger scale, without isolation/ characterization of the isonitrile and α -amino ester

intermediates, a mixture of dibromide 9 (3.64 g; 10 mmol), tetrabutylammonium hydrogen sulfate (1.53 g; 4.5 mmol), Cs₂CO₃ (3.26 g; 50 mmol), and *tert*-butyl isocyanoacetate (1.75 mL; 12 mmol) in CH₃CN (250 mL) was reacted in the same experimental conditions and workup as in Section 3.9. The crude product (containing the isonitrile **10b**) was submitted to direct acidic hydrolysis with 10 N HCl (25 drops) in CH₂Cl₂ (150 mL) and abs EtOH (75 mL), in the same experimental conditions and workup as in Section 3.11. At this stage, the crude product (containing the α -amino ester **11b**) was combined with the crude products obtained from two other similar runs in which 2.27 g (6.24 mmol) and 1.14 g (3.12 mmol) of dibromide 9 were engaged, and the mixture was purified by column chromatography on silica gel with eluant (V) to afford 1.81 g (28%) of α -amino ester **11b** containing minor impurities. This sample was treated with Fmoc–OSu (3.10 g; 9.20 mmol) in CH_2Cl_2 (400 mL) in the same experimental conditions and workup as above in Section 3.13 (a). The crude product was chromatographed on a column of silica gel with eluant (I) to afford 1.84 g (17% overall from 9) of pure 12b as a pale yellow solid. Mp 243 °C. R_f 0.63 (III). ¹H NMR (CDCl₃): δ 8.35 [s, 2H, ArH], 8.01–7.96 [m, 2H, ArH], 7.79 [br s, 2H, ArH], 7.73 [br d, J~7.5 Hz, 2H, ArH Fmoc], 7.58 [d, J=7.0 Hz, 2H, ArH Fmoc], 7.48–7.43 [m, 2H, ArH], 7.38–7.24 [m, 4H, ArH Fmoc], 5.40 [br s, 1H, NH], 4.41 [br m, 2H, CH₂ Fmoc], 4.22 [m (t-like), J~6.7 Hz, 1H, CH Fmoc], 3.72 and 3.52 [br d, $J\sim 17.3$ Hz, 2H, ArCH₂ and br d, J~17.3 Hz, 2H, ArCH₂], 1.41 [s, 9H, CH₃ O'Bu]. ¹³C NMR (CDCl₃): δ 172.3 [C=O], 155.7 [C=O Fmoc], 144.1, 141.5, 139.6, 131.8, 131.6, 128.2, 127.9, 127.3, 125.8, 125.3, 125.2, 122.6, 120.2 [C_{Ar}], 82.2 [O-C O^tBu], 67.4 [CH₂ Fmoc], 66.9 [C^α], 47.4 [CH Fmoc], 43.1 [ArCH₂], 28.0 [CH₃ O'Bu]. HRMS (FAB⁺). Calcd [M+Na]⁺ for C₃₇H₃₃NO₄Na: 578.2307. Found: 578.2313. Anal. Calcd for C₃₇H₃₃NO₄ (555.642): C, 79.97; H, 5.99; N, 2.52. Found: C, 80.45; H, 6.12; N, 2.42.

3.15. 2-*tert*-Butyloxycarbonylamino-2,3-dihydro-1*H*-cyclopenta[*b*]anthracene-2-carboxylic acid (Boc-antAib-OH 13a)

To a solution of **12a** (0.203 g; 0.5 mmol) in THF (20 mL) and MeOH (40 mL) was added a solution of 1 N NaOH (20 mL). The reaction mixture was magnetically stirred at room temperature for 24 h, cooled by addition of ice, acidified by addition of 0.5 N HCl (50 mL), and extracted with CH_2Cl_2 (2×250 mL). The CH_2Cl_2 solution was washed with H₂O (250 mL), filtered, and evaporated in vacuo. The residue was dried by repeated evaporation in vacuo at 40 °C after addition of methanol to afford 0.183 g (96%) of pure 13a as a pale yellow crystalline powder. Several similar runs gave 89–96% yields. Mp >300 °C. $R_f 0.03$ (V); 0.10 (VI). ¹H NMR (CDCl₃/CD₃OD 4:1): δ 8.22 [s, 2H, ArH], 7.87 [m, 2H, ArH], 7.68 [s, 2H, ArH], 7.34 [m, 2H, ArH], 3.66 and 3.32 [d, J~17.0 Hz, 2H and d, J~17.0 Hz, 2H, Ar-CH₂], 1.32 [s, 9H, CH₃ Boc]. ¹³C NMR (CDCl₃/ CD₃OD 4:1): δ 175.5 [C=O], 155.6 [C=O Boc], 139.0, 131.2, 131.0, 127.5, 125.0, 124.5, 121.9 [C_{Ar}], 79.5 [C–O Boc], 65.7 [C^α], 42.4 [Ar–CH₂], 27.5 [CH₃ Boc]. ESI⁺ MS m/z (relative intensity): 400.2 (100) [M+Na]⁺, 416.2 (18) [M+K]⁺, 777.4 (97) [2M+Na]⁺. HRMS (FAB⁺). Calcd [M]⁺ for C₂₃H₂₃NO₄: 377.1627. Found: 377.1629. Calcd

 $[M+H]^+$ for $C_{23}H_{24}NO_4$: 378.1705. Found: 378.1709. Anal. Calcd for $C_{23}H_{23}NO_4$ (377.422): C, 73.19; H, 6.14; N, 3.71. Found: C, 72.72; H, 6.55; N, 3.74.

3.16. 2-(9-Fluorenylmethoxycarbonylamino)-2,3-dihydro-1*H*-cyclopenta[*b*]anthracene-2-carboxylic acid (Fmoc-antAib–OH 13b)

To an ice-cold solution of 12b (0.290 g; 0.52 mmol) in CH₂Cl₂ (10 mL) was added TFA (5 mL). The solution was magnetically stirred from 0 °C to room temperature for 4 h and then evaporated in vacuo. The solid residue was repeatedly taken up in CH₂Cl₂ and the suspension evaporated in vacuo at 40 °C. The crude product was dissolved in CH₂Cl₂ (150 mL necessary) with heating, the warm solution was filtered and then concentrated to ca. 50 mL under reduced pressure at 40 °C resulting in crystallization. Cyclohexane (ca. 30 mL) was added and the mixture was concentrated again to ca. 25 mL under reduced pressure. The crystals were triturated at room temperature, filtered, abundantly washed with a solution of cyclohexane/CH₂Cl₂ ca. 95:5 (v/v), and air dried to afford 0.240 g (92%) of pure 13b as a pale yellow crystalline powder. Mp 244-246 °C. R_f 0.08 (VII), 0.42 (VIII). ¹H NMR (CDCl₃/CD₃OD 9:1 v/v): δ 8.29 [s, 2H, ArH], 7.94–7.91 [m, 2H, ArH], 7.75 [br s, 2H, ArH], 7.66 [br d, J~7.3 Hz, 2H, ArH Fmoc], 7.51 [d, J=7.3 Hz, 2H, ArH Fmoc], 7.41-7.37 [m, 2H, ArH], 7.31 [m (t-like), J~7.4 Hz, 2H, ArH Fmoc], 7.21 [m (t-like), J~7.4 Hz, 2H, ArH Fmoc], 4.33 [br m, 2H, CH₂ Fmoc], 4.15 [m (t-like), J~6.7 Hz, 1H, CH Fmoc], 3.74 and 3.46 [br d, J~16.2 Hz, 2H, ArCH₂ and br d, $J\sim$ 16.9 Hz, 2H, ArCH₂]. ¹³C NMR (CDCl₃/CD₃OD 9:1 v/v): δ 175.2 [C=O], 156.0 [C=O Fmoc], 143.6, 141.1, 139.0, 131.5, 131.3, 127.9, 127.5, 126.9, 125.5, 125.0, 122.4, 119.8 [C_{Ar}], 67.1 [C^{α}], 66.6 [CH₂ Fmoc], 47.0 [CH Fmoc], 42.7 [ArCH₂]. ESI⁺ MS m/z (relative intensity): 522.3 (100) [M+Na]⁺, 538.3 (27) $[M+K]^+$. Anal. Calcd for $C_{33}H_{25}NO_4 \cdot 0.5H_2O$ (508.546): C, 77.93; H, 5.15; N, 2.75. Found: C, 78.13; H, 5.39; N, 2.55.

3.17. Boc-antAib-L-Ala-OMe (14a)

To a suspension of 13a (0.133 g; 0.35 mmol), HCl·L-H-Ala-OMe (0.148 g; 1.06 mmol), and HOAt (0.097 g; 0.71 mmol) in THF (3 mL) and CH₂Cl₂ (3 mL) was added NMM (0.160 mL; 1.45 mmol) and then EDC (0.102 g; 0.53 mmol). The reaction mixture was stirred at room temperature for 44 h and evaporated to dryness in vacuo. The residue was solubilized in several portions of EtOAc (total of ca. 150 mL) and 0.5 N HCl (total of ca. 100 mL) with stirring, and the solutions combined and transferred into a separatory funnel. The decanted organic phase was extracted again with $0.5 \text{ N} \text{HCl}(2 \times 50 \text{ mL})$, and then H₂O(100 mL), 5% NaHCO₃ (50 mL), and H₂O (2×100 mL), dried (MgSO₄), filtered and evaporated in vacuo. The crude product was chromatographed on a preparative TLC plate of silica gel with eluant (II) to afford 0.082 g (50%) of pure 14a as a solid. Another run under similar experimental conditions and starting from 0.062 g (0.16 mmol) of 13a gave 0.047 g (62%) of pure **14a**. Mp 226–228 °C. R_f 0.40 (II). ¹H NMR (CDCl₃): δ 8.33 [s, 2H, ArH], 7.99–7.96 [m, 2H, ArH], 7.81 [s, 1H, ArH], 7.79 [s, 1H, ArH], 7.46-7.43 [m, 2H, ArH], 7.15 [br m, 1H, NH Ala], 5.22 [s, 1H, NH antAib], 4.64 [dq,

J~7.2 Hz and 7.2 Hz, 1H, CH^α Ala], 3.85 [d, J~16.7 Hz, 1H, ArCH_A antAib], 3.76 [d (partly masked), J~16.7 Hz, 1H, ArC'H_A antAib], 3.52–3.36 [br m, 2H, ArCH_B and ArC'H_B antAib], 3.74 [s, 3H, OCH₃], 1.44 [s, 9H, CH₃ Boc], 1.43 [d (partly masked), J~7.1 Hz, 3H, CH₃ Ala]. ¹³C NMR (CDCl₃): δ 173.3, 172.3 [C=O Ala and antAib], 154.9 [C=O Boc], 139.1, 131.6, 131.4, 128.0, 125.6, 125.1, 122.9, 122.8 [C_{Ar}], 80.6 [C–O Boc], 67.5 [C^α antAib], 52.4 [OCH₃], 48.3 [C^α Ala], 42.6, 42.1 [ArCH₂ antAib], 28.2 [CH₃ Boc], 18.3 [CH₃ Ala]. [α]²⁵⁸₂₉ –16.7, [α]²⁵⁷⁸ –17.3, [α]²⁵⁴⁶ –19.3, [α]²⁴³⁶ –34.0 (c 0.2; EtOAc). ESI⁺ MS *m*/z (relative intensity): 485.3 (100) [M+Na]⁺, 385.3 (40) [M+Na-Boc]⁺, 947.6 (44) [2M+Na]⁺. Anal. Calcd for C₂₇H₃₀N₂O₅·0.5H₂O (471.534): C, 68.77; H, 6.63; N, 5.94. Found: C, 69.15; H, 6.85; N, 5.93.

3.18. Fmoc-L-Ala-antAib-L-Ala-OMe (15a)

To an ice-cold solution of dipeptide 14a (0.109 g; 0.23 mmol) in CH₂Cl₂ (6 mL) was added TFA (2 mL). The solution was stirred at 0 °C for 3 h and evaporated in vacuo at 25 °C. The residue was repeatedly dissolved in CH₂Cl₂ and the solution evaporated in vacuo to yield crude TFA·H-antAib-L-Ala-OMe (not characterized). To this product was added THF (5 mL), the mixture was magnetically stirred at 0 °C for 10 min and DIEA (0.180 mL; 1.03 mmol) was added, followed by solid Fmoc-L-Ala-NCA (0.238 g; 0.71 mmol). The solution was stirred at room temperature for 66 h and evaporated in vacuo at 40 °C. The residue was dissolved in EtOAc by portions and the solutions combined and transferred to a separatory funnel. The organic solution (ca. 150 mL of EtOAc) was washed with 0.5 N HCl (2×75 mL), H₂O (2×100 mL), dried (MgSO₄), filtered, and evaporated in vacuo. The crude product was chromatographed on a preparative TLC plate of silica gel with eluant (II) (three consecutive elutions) to afford 0.128 g (83%) of pure **15a** as a solid. Mp 210–212 °C. R_f 0.15 (II); 0.55 (VII). ¹H NMR (CDCl₃): δ 8.22 [br s, 1H, ArH antAib], 8.15 [s, 1H, ArH antAib], 7.92-7.85 [m, 2H, ArH antAib], 7.73 [br m, 2H, ArH Fmoc], 7.70 [s, 1H, ArH antAib], 7.66 [s, 1H, ArH antAib], 7.43-7.34 [m, 6H, 2 ArH antAib and 4 ArH Fmoc], 7.29 [m (partly masked), 1H, NH Ala-OMe], 7.30-7.22 [m, 2H, ArH Fmoc], 6.95 [br s, 1H, NH antAib], 5.34 [d, J~6.4 Hz, 1H, NH Ala-Fmoc], 4.58 [dq, $J\sim7.2$ Hz and 7.2 Hz, 1H, CH^{α} Ala– OMe], 4.18 [m (d-like), J~6.9 Hz, 2H, CH₂O Fmoc], 4.06 [br dq, J~6.8 Hz and 6.8 Hz, 1H, CH^α Ala–Fmoc], 3.95 [br m, t-like), 1H, Ar-CH Fmoc], 3.83 [d, J~16.8 Hz, 1H, ArCH_A antAib], 3.72 [d, J~17.1 Hz, 1H, ArC'H_A antAib], 3.60 [d, $J \sim 16.9$ Hz, 1H, ArCH_B antAib], 3.47 [d, $J \sim 16.7$ Hz, 1H, ArC'H_B antAib], 3.66 [s, 3H, OCH₃], 1.38 [d, J=7.1 Hz, 3H, CH₃ Ala–OMe], 1.30 [d, J=6.9 Hz, 3H, CH₃ Ala–Fmoc]. ¹³C NMR (CDCl₃): δ 173.3, 171.8 [C=O Ala-OMe, Ala-Fmoc and antAib], 156.5 [C=O Fmoc], 143.6, 143.4, 141.1, 138.5, 131.4, 131.3, 128.7, 128.0, 127.7, 127.0, 126.9, 125.6, 125.1, 124.9, 122.8, 122.7, 119.9 [C_{Ar}], 67.8 [C^{α} antAib], 67.1 [CH₂–O Fmoc], 52.3 [OCH₃], 50.9, 48.5 [C^{α} Ala–OMe and Ala–Fmoc], 46.8 [Ar–CH Fmoc], 42.7, 41.5 [ArCH₂ and ArC'H₂ antAib], 17.8, 17.4 [CH₃ Ala–OMe and Ala–Fmoc]. $[\alpha]_{589}^{25}$ –39.4, $[\alpha]_{578}^{25} - 40.4, \ [\alpha]_{546}^{25} - 45.9, \ [\alpha]_{436}^{25} - 84.9 \ (c \ 0.11, \ CHCl_3).$ ESI⁺ MS m/z (relative intensity): 678.3 (100) [M+Na]⁺.

Anal. Calcd for $C_{40}H_{37}N_3O_6 \cdot 0.5H_2O(664.728)$: C, 72.27; H, 5.76; N, 6.32. Found: C, 72.46; H, 5.83; N, 5.91.

3.19. Synthesis of Boc-Aib-antAib-L-Ala-OMe (16a) and *cyclo*-[antAib-L-Ala] (17a)

The dipeptide 14a (0.043 g; 0.09 mmol) was N^{α} -deprotected in CH₂Cl₂ (3 mL) and TFA (1 mL) as above (Section 3.18). To the obtained crude TFA·H-antAib-L-Ala-OMe was added THF (3 mL), the mixture was magnetically stirred at 0 °C for 10 min and DIEA (0.065 mL: 0.37 mmol) was added, followed by solid Boc-Aib-NCA (0.085 g; 0.37 mmol). The solution was stirred at 60 °C for 21 h and evaporated in vacuo at 40 °C. The residue was stirred in the presence of EtOAc (10 mL). The residual solid was filtered, washed with EtOAc ($2 \times 10 \text{ mL}$), then 0.5 N HCl ($2 \times 10 \text{ mL}$), then H_2O (2×10 mL), and air dried to afford 0.015 g (48%) of pure 17a as a solid, sparingly soluble in the usual organic solvents. The filtrate (mixture of EtOAc and aqueous HCl) was diluted with more EtOAc (100 mL) and transferred to a separatory funnel. The separated organic phase was washed with 0.5 N HCl (2×50 mL), H₂O (2×100 mL), dried (MgSO₄), filtered, and evaporated in vacuo. The crude product was chromatographed on a preparative TLC plate of silica gel with eluant (II) to afford 0.007 g (14%) of pure 16a as a solid.

3.20. Boc-Aib-antAib-L-Ala-OMe (16a)

Mp 180–185 °C. R_f 0.25 (II). ¹H NMR (CDCl₃): δ 8.32 [s, 2H, ArH antAib], 7.99–7.96 [m, 2H, ArH antAib], 7.78 [m (s-like), 3H, 2 ArH antAib and NH Ala], 7.46-7.43 [m, 2H, ArH antAib], 6.65 [br s, 1H, NH antAib], 4.79 [s, 1H, NH Aib], 4.61 [dq, $J\sim7.1$ Hz and 7.1 Hz, 1H, CH^{α} Ala], 4.02 [d, J~17.0 Hz, 1H, ArCH_A antAib], 3.80 [d, J~17.0 Hz, 1H, ArC'H_A antAib], 3.54 [d, J~17.0 Hz, 1H, ArCH_B antAib], 3.23 [d, J~17.0 Hz, 1H, ArC'H_B antAib], 3.73 [s, 3H, OCH₃], 1.47 [d (partly masked), J~6.8 Hz, 3H, CH₃ Ala], 1.45 [s, 3H, CH₃ Aib], 1.42 [s, 12H, CH₃ Boc and CH₃ Aib]. ¹³C NMR (CDCl₃): δ 173.5, 172.1 [C=O Ala, antAib and Aib], 155.2 [C=O Boc], 138.7, 131.5, 128.0, 125.6, 125.1, 122.7 [C_{Ar}], 81.2 [C-O Boc], 67.6 [C^{α} antAib], 57.1 [C^{α} Aib], 52.1 [OCH₃], 48.6 [C^{α} Ala], 41.2 [ArCH₂ antAib], 28.2 [CH₃ Boc], 26.3, 22.7 [CH₃ Aib], 17.2 [CH₃ Ala]. $[\alpha]_{589}^{25}$ –35.6, $[\alpha]_{578}^{25}$ –36.9, $[\alpha]_{546}^{25}$ -41.9, $[\alpha]_{436}^{25}$ -73.3 (c 0.12, CHCl₃). ESI⁺ MS m/z (relative intensity): 570.4 (100) [M+Na]⁺. Anal. Calcd for C₃₁H₃₇N₃O₆ (547.630): C, 67.99; H, 6.81. Found: C, 68.07; H, 7.46.

3.21. cyclo-[antAib–L-Ala] (17a)

Mp >300 °C. ¹H NMR (CD₃SOCD₃): δ 8.61 [s, 1H, NH antAib], 8.45 [s, 2H, ArH antAib], 8.27 [br s, 1H, NH Ala], 8.05–8.01 [m, 2H, ArH antAib], 7.84 [s, 1H, ArH antAib], 7.83 [s, 1H, ArH antAib], 7.48–7.44 [m, 2H, ArH antAib], 4.61 [m (q-like), *J*~6.9 Hz, 1H, CH^α Ala], 3.72 [d, *J*~17.0 Hz, 1H, ArCH_A antAib], 3.61 [d, *J*~17.0 Hz, 1H, ArC'H_A antAib], 3.32 [d (partly masked), 1H, ArCH_B antAib], 3.25 [d, *J*~17.0 Hz, 1H, ArC'H_B antAib], 1.34 [d, *J*~6.9 Hz, 3H, CH₃ Ala]. ESI⁺ MS *m*/*z* (relative intensity): 331.2 (100) [M+H]⁺, 661.3

(35) $[2M+H]^+$. Anal. Calcd for $C_{21}H_{18}N_2O_2 \cdot 0.3H_2O$ (335.775): C, 75.11; H, 5.58; N, 8.34. Found: C, 75.14; H, 5.58; N, 8.47.

3.22. Ultraviolet absorption and fluorescence

The electronic absorption spectra were recorded between 300 and 400 nm using a Shimadzu model UV-2501 PC spectrophotometer. The fluorescence spectra were measured between 370 and 500 nm (upon excitation at either 359 nm or 378 nm) using a Perkin Elmer model LS-50B spectrofluorimeter. Ethanol (99.8%) was purchased from Fluka.

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